22. Heyes, J. K., and Shorland, F. B., Biochem. J., 49, 503 (1951).
23. Highlands, M. E., Licciardello, J. J., and Herb, S. F., Am. Potato J., 31, 853 (1954).
23A. Hilditch, T. P., Morton, R. A., and Riley, J. P., The Analyst, 70, 68 (1945).
23B. Hilditch, T. P., Patel, C. B., and Riley J. P., The Analyst, 76, 81 (1951).
24. Holman, R. T., and Burr, G. O., Arch. Biochem., 19, 474 (1948).
25. Jackson, J. E., Paschke, R. F., Tolberg, W. E., Boyd, H. M., and Wheeler, D. H., J. Am. Oil Chemists' Soc., 29, 229 (1952).
26. Klein, P. D., and Johnson, R. M., Arch. Biochem. and Biophys., 48, 172 (1954).
27. Klein, P. D., and Johnson, R. M., Arch. Biochem. and Biophys., 48, 380 (1954).
28. Matsuda, S., J. Soc. Chem. Ind. Japan, 45, 17 (1942).
29. Matthews, N. L., Brode, W. R., and Brown, J. B., J. Am. Chem. Soc., 63, 1064 (1941).
30. Mitchell, J. H. Jr., Kraybill, H. R., and Zscheile, F. P., Ind. Eng. Chem., Anal. Ed., 15, 1 (1943).
31. Moore, T., Biochem. J., 31, 138 (1937).
32. Mowry, D. T., Brode, W. R., and Brown, J. B., J. Biol. Chem., 142, 679 (1942).
33. Nichols, P. L. Jr., Herb, S. F., and Riemenschneider, R. W., J. Am. Chem. Soc., 73, 247 (1951).
34. Nichols, P. L. Jr., Riemenschneider, R. W., and Herb, S. F., J. Am. Oil Chemists' Soc., 27, 329 (1950).
35. O'Connell, P. W., and Daubert, B. F., Arch. Biochem., 25, 444 (1950).
36. O'Connor, R. T., Heinzelman, D. C., McKinney, R. S., and Pack, F. C., J. Am. Oil Chemists' Soc., 24, 212 (1947).
38. O'Connor, R. T., Heinzelman, D. C., McKinney, R. S., and Planck, R. W., J. Am. Oil Chemists' Soc., 24, 212 (1947).

39. Office of Rubber Reserve, Laboratory Manual, L. M. 2.3.20.2, August 1, 1945.

40. Paschke, R. F., Tolberg, W. E., and Wheeler, D. H., J. Am. Oil Chemists' Soc., 30, 97 (1953).

41. Paschke, R. F., and Wheeler, D. H., J. Am. Oil Chemists' Soc., 31, 31 (1954).

42. Pelikan, K. A., and Von Mikusch, J. D., Oil & Soap, 15, 149 (1938).

43. Planck, R. W., Pack, F. C., Heinzelman, D. C., Stansbury, M. F., and O'Connor, R. T., J. Am. Oil Chemists' Soc., 30, 598 (1953).

44. Privett, O. S., Lundberg, W. O., Khan, N. A., Tolberg, W. E., and Wheeler, D. H., J. Am. Oil Chemists' Soc., 30, 61 (1953).

45. Riemenschneider, R. W., Herb, S. F., and Nichols, P. L. Jr., J. Am. Oil Chemists' Soc., 26, 371 (1949).

46. Shorland, F. B., and Johannesson, D. L., Nature, 168, 75 (1951).

47. Swain, M. L., and Brice, B. A., J. Am. Oil Chemists' Soc., 26, 272 (1949).

48. Toyama, Y., and Shimo-oka, T., Memoirs of the Faculty of Engineering, Nagoya Univ., 5, 319 (1953).

50. Toyama, Y., and Shimo-oka, T., Memoirs of the Faculty of Engineering, Nagoya Univ., 5, 323 (1953).

51. Toyama, Y., and Tsuchiya, T., Bull. Chem. Soc. Japan, 10, 192 (1935).

52. Toyama, Y., and Yamamoto, T., Memoirs of the Faculty of Engineering, Nagoya Univ., 5, 114 (1953).

53. Von Mikusch, J. D., J. Am. Oil Chemists' Soc., 29, 114 (1952).

54. Widmer, C. Jr., and Holman, R. T., Arch. Biochem., 202, 417 (1953).

56. Wiese, H. F., and Hansen, A. E., J. Biol. Chem., 202, 417 (1953).

[Received September 27, 1954]

Comparative Antioxidant Potency of 2,4,5-Trihydroxyphenones

M B KNOWLES, ALAN BELL, CLARENCE E. THOLSTRUP, and HERMAN S. PRIDGEN, Research Laboratories, Tennessee Eastman Company, Division of Eastman Kodak Company, Kingsport, Tennessee

This report is concerned with the results of testing 2,4,5-trihydroxyphenones as antioxidants for fats, paraffin wax, and mineral oil. The use of antioxidants for stabilizing various organic materials has grown fairly rapidly the last few years. More and more specific applications are being discovered as more antioxidant compositions are produced in the laboratory and become available for testing. Included in the current applications are eracked gasolines, leaded aviation gasoline, turbine oil, jet fuel, certain vitamins (such as carotene), rubber, paraffin wax, polymeric materials (such as polyethylene), and fats and oils. The increasing use of fats in cattle and poultry feeds will undoubtedly point out the need for antioxidants in these materials.

Antioxidants in edible fats serve two purposes: a) they prolong storage life by preventing rancidity development and b) they increase shelf life of foods cooked with or containing the stabilized fat. At present there are several compounds which are approved by the Food and Drug Administration for use in foods. These compounds are butylated hydroxyanisole (BHA), propyl gallate (PG), citric acid (CA), butylated hydroxytoluene (BHT), nordihydroguaiaretic acid (NDGA), guaiac acid, tocopherols, lecithin, phosphoric acid, 3,3'-thiodipropionic acid, dilauryl and distearyl esters of 3,3'-thiodipropionic acid, and ascorbic acid.

It might be expected that antioxidants for edible fats should also be antioxidants for non-edible fats. This is indeed true, but in many cases the non-edible varieties are more difficult to stabilize.

A series of compounds was tested as antioxidants for several different media by means of the Swift Active Oxygen Method (AOM). These compounds are represented by the formula

in which R is hydrogen, alkyl, cycloalkyl, aralkyl, or aryl. These compounds, in most AOM tests, are highly potent antioxidants. They are outstandingly good in lard, paraffin wax, and mineral oil and have some degree of potency in the various oils tested.

Discussion

Table I presents some AOM data for lard stabilized with the 2,4,5-trihydroxyphenones and also shows corresponding data for BHA, BHT, and PG. The data show that 2,4,5-trihydroxyphenones have very high potencies in lard and that the potency decreases with increasing molecular weight. This is shown graphically in Figure 1, in which AOM values are plotted against the number of carbon atoms in the R group. It is noted that the two compounds with branched R groups do not fall on the same line as that for the compounds with straight-chain R groups.

In Figure 2 AOM values for lard are plotted against antioxidant concentration. This plot shows that the AOM values for the 2,4,5-trihydroxyphenones are more nearly proportional to concentration than is the case with BHA, BHT, or PG.

In Table II AOM stability data for some of the 2,4,5-trihydroxyphenones in certain edible oils are given. In all cases the AOM life was increased. In peanut oil the effectiveness was most pronounced; the increase approached four-fold in this particular sample. In cottonseed oil the life increase was nearer

TABLE 1
Results of Tests with Lard

Antioxidant	Stability, Hrs. (20 meq. peroxide)				
Compound	Concn., % by wt.	Sample A	Sample B	Sample C	
None		13	13	12	
BHA	0.02	31	35	31	
PG	0.02	89	127	92	
BHT	0.02	44	54	47	
R Group of Phenone					
Hydrogen	0.01	96			
	0.02	193			
Methyl	0.01	128			
	0.02	205			
Propyl	0.005	67			
	0.010	122			
	0.020	163			
Isopropyl	0.02	167			
Propenyl	0.01		72		
Amyl	0.02			106	
3-Amyl	0.02			75	
Heptyl	0.005	l	71		
	0.010		88		
	0.020	160	168		
3-Heptyl	0.02		100	69	
Nonyl	0.02			106	
Undecyl	0.02			106	
Heptadecyl	0.01		74		
Benzyl	0.02		l	120	
Cyclohexyl	0.02		l	87	
Phenyl	0.01	100			
	0.02	172	l	1	
2-Furyl	0.005	64			
	0.010	84			
	0.020	127			

two-fold. The lack of sufficient data on corn oil prohibits the drawing of any conclusions.

Some AOM data for paraffin wax and mineral oil are presented in Table III. It should be noted that data are shown for both 100°C. and 150°C. The higher temperature is more convenient in that the time required for the test is much shorter. The 2,4,5-trihydroxyphenones that were tested appeared to be quite effective in preventing peroxide formation under the test conditions.

In Table IV AOM data for two of the 2,4,5-trihy-droxyphenones in castor, palm, sperm, and sesame oils are given. The two phenones were fairly effective in all of these oils and increased AOM life considerably. None of the antioxidants shown were very potent in palm or sperm oils.

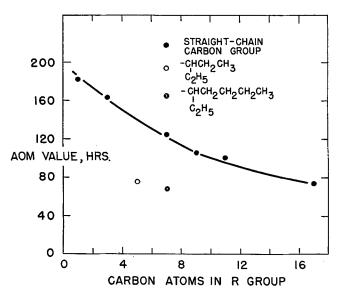


Fig. 1. Effect of R group on AOM keeping quality of lard stabilized with 2,4,5-trihydroxyphenones.

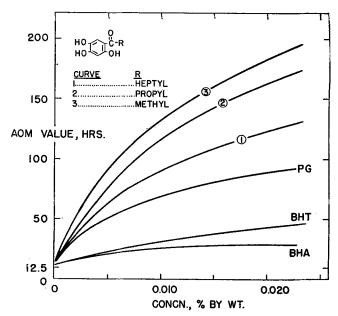


Fig. 2. Effect of antioxidant concentration on AOM keeping quality of lard.

AOM data for tallows and greases stabilized with various antioxidants are shown in Table V. All of the compounds were effective in tallows, but none were outstandingly good in greases except when used with citric acid. By using citric acid as a

TABLE II
Results of Tests with Cottonseed, Peanut, and Corn Oils

	Stability, Hrs. (70 meq. peroxide)					
Antioxidant ^a	Cotto	nseed				
	Sample A	Sample B	Peanut	Corn		
None	11	11	12	6		
BHA	11	11	12	6		
PG	27	53	48	11		
BHT	11	11	25	11		
R Group of Phenone						
Hydrogen	30		47	11		
Methyl			48	15		
Propyl			62			
Isopropyl		23	55			
Propenyl		18				
Heptyl		20	52			
Heptadecyl		1 17	45			
Phenyl			52			

^a Concn. = 0.02% by wt.

TABLE III

Results of Tests with Mineral Oil and Paraffin Wax at 100°C. and 150°C.

Antioxidant		Stability, Hrs. (50 meq. peroxide)					
		Pa	35:1				
Compound	Concn.,	Sample A Samp		le B	Mineral oilb		
	% by wt.	150°C.	100°C.	150°C.	150°C.		
None		1.7	54	1	0.5		
BHA	0.02	18		••••			
PG	0.01		670	17	17		
BHT	0.01		135	5	6		
R Group of Phenone	1						
Methyl	0.01	29	475	15	17		
Propyl	0.01	62	>1,000	29	21		
Heptyl				26	20		
Isopropyl	0.01			21			
Nonyl	0.01			21			
Undecyl	0.01			23	****		
Phenyl				25			

^a Aristowax, amp. 130/134. ^b Primol D, flash point 433°F.(223°C.).

TABLE IV Results of Tests with Castor, Palm, Sperm, and Sesame Oils

Antioxidant ^a	Stability, Hrs. (70 meq. peroxide)					
	Castor	Palm	Sperm	Sesame		
None	91	3.0	1.0	4.5		
PG	210	3.5	1.0	16.0		
BHT	97	4.0	2.0	5.5		
BHA R Group of Phenone	95	4.0	3.5	4.5		
Methyl	139	4.5	3.0	25.0		
Propyl	117	4.0	2.5	12.5		

TABLE V Results of Tests with Tallows and Greases

Antioxidant		Stability, Hrs. (20 meq. peroxide)						
		Greases						
	Tallows		Choice white		Yellow			
	Fancy	Choice	Prime	Aa	Bb	Aa	\mathbf{B}_{p}	
None	28	10	23	0.5	1.0	2.0	30.0	
BHA	170	> 260	83	0.5	46.0	5.0	74.0	
PG	89	>260	35	0.5	63.0	3.5	57.0	
BHT	95	197	43	0.5	14.0	3.0	32.0	
R Group of Phenone								
Methyl	100	> 260	31	0.5	10.0	5.5	60.0	
Propyl	96	> 260	40	0.5	5.0	6.0	71.0	

a 0.01% antioxidant.
b 0.01% antioxidant plus 0.05% citric acid.

metal scavenger, or synergist, the greases were stabilized satisfactorily.

Conclusions

On the basis of AOM tests run to date it appears that 2,4,5-trihydroxyphenones are quite potent antioxidants for many applications. Some of the lower members of the series, such as 2,4,5-trihydroxybutyrophenone, are outstandingly effective in lard, paraffin wax, mineral oil, and peanut oil. In other media, such as cottonseed oil, corn oil, castor oil, sesame oil, and tallows, they are less effective but still fairly active. In greases, palm oil, and sperm oil, they are only moderately effective. Over-all the 2,4,5-trihydroxyphenones compare quite favorably with current commercial products. It would appear that as more testing is done, more applications for these compounds may be found.

Subacute toxicity studies made with 2,4,5-trihydroxyacetophenone and 2,4,5-trihydroxybutyrophenone indicate that they are of a low order of toxicity. Long-term feeding tests on rats and dogs are underway with the butyrophenone, and after four months

there are no apparent ill effects.

[Received October 14, 1954]

Present Status of the Filtration-Extraction Process for Cottonseed

J. J. SPADARO, H. L. E. VIX, H. K. GARDNER, E. L. D'AQUIN, P. H. EAVES, and E. A. GASTROCK, Southern Regional Research Laboratory, New Orleans, Louisiana

EVELOPMENT of the various phases of the filtration-extraction process for cottonseed has been reported in previous papers (1, 2, 3, 4). Since the publication of these papers many important improvements or "process refinements" have been made, and some of these have been applied commercially in the first large-scale plant which was installed in Mississippi. Many of these improvements have been developed on a cooperative basis with the Mississippi Cottonseed Products Company³ and the Lukenweld Division of Lukens Steel Company.³

Improvements or changes in the processing procedures and operating features, which depict the present status of the development of the process for cottonseed, involve the following phases of the process: conditioning of the meats or flakes prior to cooking: extraction temperature; effect of screw conveying both flakes and cooked meats; effect of an open-weave type filter medium; effect of recycling the initial concentrated miscella filtrate; effect of reforming cooked material; effect of cake thickness; and product quality. The following brief discussion of the filtration-extraction process is required better to understand the application of the process refinements.

Essential steps of the process are shown in Figure

1. Prepared cottonseed flakes are cooked in conventional four-to-six-high stack cookers, using lower temperatures and shorter cooking time than normally used for hydraulic cooking. Moisture content of the material is maintained throughout the cooking period at levels higher than for hydraulic cooking. The cooked material is "crisped" by evaporative cooling and by reducing the size of any large agglomerates formed during the cooking operation. The conditioned material is mixed with one of the miscella filtrates to form a slurry which is held for 25 to 40 min. The slurry is filtered, and the resulting cake countercurrently washed three times on a horizontal rotary vacuum filter. Total time on the filter is only 2 to 4 min. The solvent damp extracted meal (marc) is discharged to a conventional desolventizer for meal and solvent recovery. The concentrated miscella is pumped to the oil recovery system for separation of oil and solvent.

Conditioning of Meats or Flakes. Cottonseed used in the earlier pilot-plant runs had a normal average moisture content of 9 to 11%. The moisture of the resulting flakes was increased to the required 16 to 22% in the first or second ring of the cooker. In some of the recent runs the use of cottonseed having a moisture content of 6 to 8% showed the preferability of altering the method of adding the water. The drier meats from this seed tended to produce more fines when flaked to 0.008 to 0.010 in.; and the resulting flakes required the addition of more water to at-

¹ Presented at the 45th annual meeting of the American Oil Chemists' Society, San Antonio, Tex., April 11-14, 1954.

² One of the laboratories of the Southern Utilization Research Branch, Agricultural Research Service, U. S. Department of Agriculture.

³ The Agricultural Research Service has memorandum of understanding with this Company.